What Choline Metabolism Can Tell Us About the Underlying Mechanisms of Fetal Alcohol Spectrum Disorders

Steven H. Zeisel

Received: 16 November 2010 / Accepted: 5 January 2011 / Published online: 25 January 2011 © Springer Science+Business Media, LLC 2011

Abstract The consequences of fetal exposure to alcohol are very diverse and the likely molecular mechanisms involved must be able to explain how so many developmental processes could go awry. If pregnant rat dams are fed alcohol, their pups develop abnormalities characteristic of fetal alcohol spectrum disorders (FASD), but if these rat dams were also treated with choline, the effects from ethanol were attenuated in their pups. Choline is an essential nutrient in humans, and is an important methyl group donor. Alcohol exposure disturbs the metabolism of choline and other methyl donors. Availability of choline during gestation directly influences epigenetic marks on DNA and histones, and alters gene expression needed for normal neural and endothelial progenitor cell proliferation. Maternal diets low in choline alter development of the mouse hippocampus, and decrement memory for life. Women eating lowcholine diets have an increased risk of having an infant with a neural tube or orofacial cleft birth defect. Thus, the varied effects of choline could affect the expression of FASD, and studies on choline might shed some light on the underlying molecular mechanisms responsible for FASD.

Keywords Choline · Epigenetics · Methyl donor · Alcohol · Single nucleotide polymorphism · Nutrition

S. H. Zeisel (⊠)

Department of Nutrition, UNC Nutrition Research Institute at Kannapolis, University of North Carolina at Chapel Hill, 500 Laureate Way, Room 2218, Kannapolis, NC 28081, USA

e-mail: steven zeisel@unc.edu

Introduction

Exposure of germ cells, fetus, and young infant to alcohol can cause abnormalities in behavior and organ structures that range from barely detectable, to birth defects, to fetal loss [1]. Fetal alcohol spectrum disorders (FASD) include growth retardation, distinctive facial anomalies, cardiac defects, and altered brain function; less commonly, FASD are associated with skeletal, ocular, vestibular, hepatic, skin, and immune defects [2]. Thus, the consequences of fetal exposure to alcohol are very diverse and the likely molecular mechanisms involved must be able to explain how so many developmental pathways could go awry. Though there are likely many varied mechanisms of damage, Haycock [3] recently hypothesized that epigenetic factors are among the important mechanisms underlying FASD based on the important role of epigenetic mechanisms in central nervous system development.

Though our genetic code is spelled out at the time of conception, humans retain some level of flexibility about which of our genes is active or suppressed. This is accomplished by an epigenetic code which is transmitted by DNA methylation, covalent modifications of histones and chromatin, and by RNA interference [4]. The pattern of DNA methylation is first established during gastrulation [5] but can be changed during fetal development and early life. Haycock argues that epigenetic perturbations could cause the varied sequelae observed after exposure to alcohol [3]. This concept is strengthened by evidence that chronic exposure to alcohol perturbs one-carbon metabolism, thereby altering the key substrates needed to methylate DNA and histones [6].

A parallel line of investigation seems to lead to the same conclusion—that FASD may be caused by epigenetic perturbations at the time that progenitor cells are dividing



and differentiating to form organs. Children with FASD have a perturbation in one-carbon metabolism that is reflected in a decreased choline/creatine ratio in brain (left striatum) [7]. In 2009, Thomas and colleagues [8] reported that when pregnant rat dams were exposed to ethanol during gestational days 5-20, their offspring had reduced birth weight and brain weight, delays in eye opening and incisor emergence, and alterations in the development of the righting reflex, geotactic reflex, cliff avoidance, reflex suspension, and hindlimb coordination. However, if rat dams were also treated with choline, the effects from ethanol were attenuated (on birth and brain weight, incisor emergence, and most behavioral measures) in their pups. In fact, behavioral performance of ethanolexposed pups treated with choline did not differ from that of controls. This effect from choline occurred without changing alcohol exposure, as blood alcohol levels were not changed by choline [8]. If children with FASD have some perturbation in choline metabolism, and choline supplementation attenuates the effects of alcohol exposure, then perhaps examining choline-related mechanisms will be of use in understanding the molecular etiology of FASD (Fig. 1).

Choline and Fetal Development

Choline is an essential nutrient for humans [9], and it is needed for biological membrane formation, for methylation, and for acetylcholine biosynthesis [10]. Choline is found in a variety of foods ([11]; and see http://www.ars. usda.gov/services/docs.htm?docid=6232; last accessed 11 Nov 2010), but the richest food sources of choline (eggs and liver) are avoided by many women. In fact, in the USA, most women do not eat diets delivering the recommended intake of choline [12]. Newborns have higher tissue choline concentrations than do their mothers at term (e.g., 10-15fold higher in rats) [13, 14] because the placenta effectively takes choline from mother and delivers it to the fetus [15– 17]. Later on, mother keeps supplying the baby with choline, as the mammary gland also delivers choline from mother to baby in breast milk [18, 19]. Finally, pregnant women may have a higher capacity to form phosphatidylcholine (a source of choline) from precursors in liver because the gene PEMT, that encodes the enzyme phosphatidylethanolamine-N-methyltransferase that catalyzes this process, is induced by estrogen [20]. Thus, there are many physiological mechanisms that permit women to

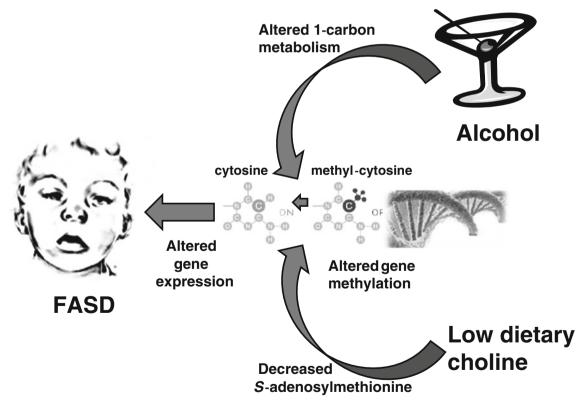


Fig. 1 Potential shared mechanisms. Diets low in choline, or genetic variations that increase the dietary demand for choline, can decrease DNA methylation (converting methyl-cytosines to cytosines in genes) and this increases gene transcription, changes progenitor cell proliferation and differentiation, resulting in birth defects and

abnormal brain development. It is possible that similar mechanisms explain how alcohol exposure causes fetal alcohol spectrum disorders (*FASD*). Alcohol exposure during pregnancy perturbs one-carbon metabolism, decreasing the use of folate and increasing the use of choline as a methyl donor



deliver large quantities of choline to the growing fetus and infant and, for this to have evolved choline must play some important role in development.

Rodent dams fed diets low in choline during days 11–17 of gestation had pups with diminished neural progenitor cell proliferation in fetal hippocampus at the same time that there was increased apoptosis in these cells [21, 22]. In electrophysiologic studies, an insensitivity to long-term potentiation (LTP) was measured when these pups were adult animals [23], and in cognitive studies, they had decremented visuospatial and auditory memory as adults [24–27]. Conversely, rodent dams fed diets supplemented with choline (about four times the normal chow levels) during days 11-17 of gestation had pups with increased sensitivity to LTP when they were adult animals [23], and enhanced visuospatial and auditory memory as adults [24– 27]. During gestation, maternal diets low in choline were also associated with increased neural tube closure defects and orofacial defects in rodent and human fetuses [28–31]. Apparently, choline availability is important for normal fetal development throughout gestation.

Mechanisms for the Actions of Choline

One of the likely mechanisms for these effects of choline on fetal development is epigenetically mediated. Choline is a major source of methyl groups [10], and, as discussed earlier, methylation of DNA and histones are important components of the epigenetic code [4]. Thus, DNA methylation is altered by availability of dietary choline [32]. DNA can be methylated on cytosines, and when this nucleotide is methylated in a specific location with the promoter of a gene, methyl-binding proteins are attracted, forming a complex that prevents access by transcription factors needed to activate expression of the gene [4]. Thus, usually methylation inhibits gene expression and hypomethylation increases gene expression. In the rodent experiments on dietary choline and memory that were described earlier, dams fed diets low in choline during days 11–17 of gestation had pups with diminished DNA and histone methylation in genes of the hippocampus [33, 34]. Specifically, a gene (cdkn3) that normally inhibits cell cycling, was hypomethylated and overexpressed in the prime germinal zone of the dentate gyrus of the fetal hippocampus [34, 35]. The product of this gene, kinase-associated phosphatase (Kap) protein levels were increased and this subsequently activated the retinoblastoma protein (Rb) pathway that inhibits cyclin-dependent kinase [34, 35].

In mice, maternal diets low in choline during pregnancy also altered histone methylation in fetal neural progenitor cells in the areas of the hippocampus where neurogenesis was occurring (supraventricular and ventricular zones; [33]). Transcriptional repressor neuron-restrictive silencing factor (REST) binding to gene promoter regions mediates the inhibition of expression of numerous neuronal genes and is a very important regulator of brain development [36]. REST recruits a group of co-repressors (histone deacety-lase, methyl CpG binding protein 2 [37], and G9a histone methyltransferase [38]) to the repressor element 1 (RE1) in the promoter of various genes, resulting in inhibition of neuronal gene expression. The binding of REST at the RE1 site is facilitated by a specific histone methylation pattern [39]. When neural progenitor cells were exposed to a low-choline environment in vitro, there was diminished REST binding to RE1 [33]. Thus, availability of choline modulated histone methylation and thereby gene expression in fetal neural progenitor cells.

Choline effects are not limited to neural progenitor cells. We examined whether maternal dietary choline modulates angiogenesis in fetal brain [40]. In fetuses from mouse dams fed a low-choline diet during pregnancy, proliferation of endothelial cells in hippocampus was decreased because endothelial progenitor cells differentiated prematurely and stopped dividing [40]. These changes were associated with >25% decrease in the number of blood vessels in fetal hippocampus in the lowcholine group (p < 0.01 vs. control). In developing brain, it is important to maintain a balance between angiogenesis and neurogenesis, and this is accomplished by careful modulation by local cues (growth factors, extracellular matrix) [41-45] that include vascular endothelial growth factors (VEGF) and their receptors and by angiopoietin (ANGPT)/endothelial receptor tyrosine kinase signaling (Tie-2) [46-48]. Cytosines in the promoter regions of vascular endothelial factor C (VEGFC) and angiopoietin 2 (ANGPT2) were hypomethylated in neural progenitor cells exposed to choline compared to controls, providing an explanation for why these genes were over expressed [40]. Expression of genes for these angiogenic signals was increased in fetal hippocampus in the low-choline group (VEGFC, 2.0-fold, p < 0.01 vs. control and ANGPT2, 2.1fold, (p < 0.01 vs. control) [40]. Thus, maternal dietary choline intake and epigenetic changes also alter angiogenesis in the developing fetal hippocampus.

Shared Mechanisms for Effects of Choline and Alcohol?

Several types of tissues (nerves and blood vessels; perhaps others) are capable of changing epigenetic marks when presented with a low-choline environment. Choline availability during gestation in rodents modifies DNA methylation, DNA methyltransferase expression, histone H3 methylation, and histone methyltransferase expression in fetal tissues [33, 34, 49, 50]. However, these effects may occur only during critical windows of time during gestation



when progenitor cells are exposed to proliferationpromoting signals and before they differentiate. Hippocampal neural progenitor cell proliferation occurs from day 11-17 of mouse gestation, and that period is when choline exerts its effects. Restoration of choline later in development may not correct the epigenetic marks. Though not yet proven, it is likely that other types of progenitor cells, developing at different times during gestation, are also sensitive to choline. This could explain choline effects on neural tube and cleft palate defects. Exposure to alcohol also changes epigenetic marks. Maternal exposure to ethanol during pregnancy resulted in genome-wide hypomethylation in fetuses [51], and to differential modulation of H19 DNA methylation (controls an important growth factor gene) in the paternal and maternal alleles in the placenta [52]. Paternal alcohol exposure altered methylation of imprinted genes in the male gamete and decreased cytosine methyltransferase mRNA levels in paternal sperm [53, 54]. Thus, alcohol (and perhaps choline) might exert different effects on the epigenetic regulation of gene expression in the gametes, fetus, and placenta.

Alcohol and Choline Metabolism

Choline, folate, and methionine metabolism are highly inter-related, and these pathways intersect at the formation of methionine from homocysteine [10]. Homocysteine can be methylated using methyltetrahydrofolate or the choline metabolite betaine as the methyl group donor [10]. Manipulations that reduce the availability of methyl groups from folate increase the demand for choline and vice versa [55-57]. In rats, ethanol feeding inhibits the use of folate methyl groups to make methionine (methionine synthetase activity inhibited) and increases the use of betaine to make methionine (activity of the enzyme betaine-homocysteine methyl transferase is increased) [58]. Acute ingestion of alcohol in humans lowers brain concentrations of choline as measured by magnetic resonance spectroscopy (the choline/ creatinine rations measured in such imaging likely measures a mixture of choline-containing compounds in brain) [59]. In the micropig, transcription of genes and activity for several other enzymes in one-carbon metabolism is altered by exposure to alcohol (i.e., methyltetrahydrofolate reductase, methionine adenosyltransferase 1A, glycine Nmethyltransferase, S-adenosylhomocysteine hydrolase) [60]. In alcoholic liver disease, methionine metabolism is impaired, and S-adenosylmethionine (formed from methionine) concentrations in liver are decreased [61]. Sadenosylmethionine is the methyl donor needed for methylation of DNA and histones. Alcohol exposure also diminishes the availability of methyltetrahydrofolate, thereby increasing the demand for choline. Diets of alcoholics are especially deficient in folate [62]. Very low dietary folate intake (<180 µg per day) was 2.5-fold more common among women who drank 30 g alcohol regularly [63]. Heavy alcohol users malabsorb folate [64] and increase the loss of folate in the urine through a reduction in renal tubular reabsorption [65]. It is interesting that, in some animal models, dietary betaine supplementation attenuates several of the metabolic abnormalities associated with alcohol, correcting the abnormal hepatic *S*-adenosylmethionine and homocysteine concentrations and mitigating some of the damage to the liver [66, 67].

Could Mechanisms Related to Epigenetics and Choline be the Cause of FASD?

The consequences of fetal exposure to alcohol include growth retardation; distinctive facial anomalies; cardiac defects; altered brain function; and skeletal, ocular, vestibular, hepatic, skin, and immune defects [2]. Choline deficiency during gestation in animal models is associated with growth retardation, altered brain function, skeletal abnormalities, hepatic and muscle damage, as well as orofacial and neural tube birth defects [10]. Decreased choline availability to the fetus decreases hippocampal neurogenesis and increases apoptosis [68]. Exposure of the fetus to alcohol also decreases hippocampal neurogenesis and decreases cell survival [69, 70], resulting in reduced numbers of hippocampal pyramidal cells [71]. Though there are differences in the genes and tissues studied in both models, both choline deficiency and ethanol alter genes of cell cycling by altering DNA methylation of these genes [35, 72]. Prenatal alcohol exposure perturbs behavioral development leading to hyperactivity, motor incoordination, alterations in social processing, and to deficits in cognitive functioning, and prenatal supplementation with choline prevents or mitigates these effects [73]. Could there be shared molecular mechanisms that explain how so many developmental pathways could be altered?

Choline and epigenetics can also influence brain function. As discussed earlier, choline availability to pregnant rodents alters memory function in their juvenile, adult, and aged offspring [74]. In a mouse model of Down Syndrome, mice born to mothers supplemented with choline during the perinatal period had significant improvements in cognitive function and emotion regulation [75]. Epigenetic mechanisms underlie a number of syndromes in which brain function abnormalities are apparent, including Rett syndrome, an autism spectrum disorder that is characterized by cognitive impairments [76]. This syndrome results from mutations in the methyl DNA-binding protein MeCP2 [77]. Also, there are data suggesting that epigenetic factors are associated with hyperactivity in children [78] and certain psychiatric disorders [79]. As a whole, the available data suggests that the underlying mechanism responsible for the



effects of choline could also explain how many developmental pathways could go awry in FASD.

Clues that Might be of Interest for Future Studies in FASD

Humans have widely varying dietary requirements for choline, in part explained by genetic variation. As discussed earlier, several metabolic pathways influence how much choline is required from diet, and single nucleotide polymorphisms (SNPs) in specific genes influence the efficiency of these pathways. Specifically, some polymorphisms in the folate pathways limit the availability of methyltetrahydrofolate and thereby increase use of choline as a methyl donor; polymorphisms in the PEMT gene alter endogenous synthesis of choline; and polymorphisms in other genes of choline metabolism influence dietary requirements by changing the utilization of choline moiety [80, 81]. At this time we do not know whether maternal or fetal SNPs (or some combination of both) are most important in setting demand for choline. The fact that these SNPs are very common suggests that they could be overcome by high choline in the diet, perhaps because humans can eat choline to overcome the inefficiencies associated with the SNPs. If FASD shares some common mechanism with the effects of choline, then it is possible that people with SNP-induced metabolic inefficiencies who also eat a low-choline diet are at greater risk for FASD.

Acknowledgements This work was supported by grants from the NIH (DK55865, DK56350).

References

- Hoyme HE, May PA, Kalberg WO, Kodituwakku P, Gossage JP, Trujillo PM, Buckley DG, Miller JH, Aragon AS, Khaole N, Viljoen DL, Jones KL, Robinson LK (2005) A practical clinical approach to diagnosis of fetal alcohol spectrum disorders: clarification of the 1996 institute of medicine criteria. Pediatrics 115:39–47
- Chaudhuri JD (2000) Alcohol and the developing fetus—a review. Med Sci Monit 6:1031–1041
- Haycock PC (2009) Fetal alcohol spectrum disorders: the epigenetic perspective. Biol Reprod 81:607–617
- Zeisel SH (2009) Epigenetic mechanisms for nutrition determinants of later health outcomes. Am J Clin Nutr 89:1488S

 1493S
- Reik W, Dean W, Walter J (2001) Epigenetic reprogramming in mammalian development. Science 293:1089–1093
- Halsted CH, Villanueva JA, Devlin AM, Niemela O, Parkkila S, Garrow TA, Wallock LM, Shigenaga MK, Melnyk S, James SJ (2002) Folate deficiency disturbs hepatic methionine metabolism and promotes liver injury in the ethanol-fed micropig. Proc Natl Acad Sci USA 99:10072–10077
- Goncalves Rde C, Vasconcelos MM, Faleiros LO, Cruz LC Jr, Domingues RC, Brito AR, Werner J Jr, Herdy GV (2009) Proton magnetic resonance spectroscopy in children with fetal alcohol spectrum disorders. Arq Neuropsiquiatr 67:254–261

- Thomas JD, Abou EJ, Dominguez HD (2009) Prenatal choline supplementation mitigates the adverse effects of prenatal alcohol exposure on development in rats. Neurotoxicol Teratol 31:303– 311
- Institute of Medicine, and National Academy of Sciences USA (1998) Choline. In: Dietary reference intakes for folate, thiamin, riboflavin, niacin, vitamin B12, panthothenic acid, biotin, and choline Vol. 1. National Academy Press, Washington DC, pp 390– 422
- Zeisel SH (2006) Choline: critical role during fetal development and dietary requirements in adults. Annu Rev Nutr 26:229–250
- Zeisel SH, Mar MH, Howe JC, Holden JM (2003) Concentrations of choline-containing compounds and betaine in common foods. J Nutr 133:1302–1307
- Jensen HH, Batres-Marquez SP, Carriquiry A, Schalinske KL (2007) Choline in the diets of the U.S. population: NHANES, 2003–2004. FASEB J 21:lb219
- Zeisel SH, Wurtman RJ (1981) Developmental changes in rat blood choline concentration. Biochem J 198:565–570
- Ozarda Ilcol Y, Uncu G, Ulus IH (2002) Free and phospholipidbound choline concentrations in serum during pregnancy, after delivery and in newborns. Arch Physiol Biochem 110:393–399
- Sweiry JH, Page KR, Dacke CG, Abramovich DR, Yudilevich DL (1986) Evidence of saturable uptake mechanisms at maternal and fetal sides of the perfused human placenta by rapid paired-tracer dilution: studies with calcium and choline. J Devel Physiol 8:435– 445
- Banns HE, Stephens NL (1974) Proceedings: investigation of choline acetylase from various tissues by use of ionizing radiation. J Physiol 242:16P–18P
- 17. Yudilevich DL, Sweiry JH (1985) Membrane carriers and receptors at maternal and fetal sides of the placenta by single circulation paired-tracer dilution: evidence for a choline transport system. Contrib Gynecol Obstet 13:158–161
- Zeisel SH, Char D, Sheard NF (1986) Choline, phosphatidylcholine and sphingomyelin in human and bovine milk and infant formulas. J Nutr 116:50–58
- Holmes-McNary MQ, Cheng WL, Mar MH, Fussell S, Zeisel SH (1996) Choline and choline esters in human and rat milk and in infant formulas. Am J Clin Nutr 64:572–576
- Resseguie M, Song J, Niculescu MD, da Costa KA, Randall TA, Zeisel SH (2007) Phosphatidylethanolamine N-methyltransferase (PEMT) gene expression is induced by estrogen in human and mouse primary hepatocytes. FASEB J 21:2622–2632
- Albright CD, Tsai AY, Friedrich CB, Mar MH, Zeisel SH (1999) Choline availability alters embryonic development of the hippocampus and septum in the rat. Brain Res 113:13–20
- Albright CD, Friedrich CB, Brown EC, Mar MH, Zeisel SH (1999) Maternal dietary choline availability alters mitosis, apoptosis and the localization of TOAD-64 protein in the developing fetal rat septum. Brain Res 115:123–129
- Jones JP, Meck W, Williams CL, Wilson WA, Swartzwelder HS (1999) Choline availability to the developing rat fetus alters adult hippocampal long-term potentiation. Brain Res 118:159– 167
- 24. Cheng RK, MacDonald CJ, Williams CL, Meck WH (2008) Prenatal choline supplementation alters the timing, emotion, and memory performance (TEMP) of adult male and female rats as indexed by differential reinforcement of low-rate schedule behavior. Lear Mem 15:153–162
- Meck W, Williams C (1997) Perinatal choline supplementation increases the threshold for chunking in spatial memory. Neuro-Report 8:3053–3059
- Meck WH, Smith RA, Williams CL (1988) Pre- and postnatal choline supplementation produces long-term facilitation of spatial memory. Dev Psychobiol 21:339–353



- Meck WH, Williams CL (1999) Choline supplementation during prenatal development reduces proactive interference in spatial memory. Brain Res 118:51–59
- Fisher MC, Zeisel SH, Mar MH, Sadler TW (2001) Inhibitors of choline uptake and metabolism cause developmental abnormalities in neurolating mouse embryos. Teratology 64:114–122
- Fisher MC, Zeisel SH, Mar MH, Sadler TW (2002) Perturbations in choline metabolism cause neural tube defects in mouse embryos in vitro. FASEB J 16:619–621
- Blom HJ, Shaw GM, den Heijer M, Finnell RH (2006) Neural tube defects and folate: case far from closed. Nat rev 7:724–731
- Shaw GM, Carmichael SL, Yang W, Selvin S, Schaffer DM (2004) Periconceptional dietary intake of choline and betaine and neural tube defects in offspring. Am J Epidemiol 160:102–109
- Alonso-Aperte E, Varela-Moreiras G (1996) Brain folates and DNA methylation in rats fed a choline deficient diet or treated with low doses of methotrexate. Int J Vitam Nutr Res 66:232–236
- Mehedint MG, Niculescu MD, Craciunescu CN, Zeisel SH (2010) Choline deficiency alters global histone methylation and epigenetic marking at the Re1 site of the calbindin 1 gene. FASEB J 24:184–195
- Niculescu MD, Craciunescu CN, Zeisel SH (2006) Dietary choline deficiency alters global and gene-specific DNA methylation in the developing hippocampus of mouse fetal brains. FASEB J 20:43–49
- 35. Niculescu MD, Yamamuro Y, Zeisel SH (2004) Choline availability modulates human neuroblastoma cell proliferation and alters the methylation of the promoter region of the cyclindependent kinase inhibitor 3 gene. J Neurochem 89:1252–1259
- 36. Chong JA, Tapia-Ramirez J, Kim S, Toledo-Aral JJ, Zheng Y, Boutros MC, Altshuller YM, Frohman MA, Kraner SD, Mandel G (1995) REST: a mammalian silencer protein that restricts sodium channel gene expression to neurons. Cell 80:949–957
- Ballas N, Grunseich C, Lu DD, Speh JC, Mandel G (2005) REST and its corepressors mediate plasticity of neuronal gene chromatin throughout neurogenesis. Cell 121:645–657
- Roopra A, Qazi R, Schoenike B, Daley TJ, Morrison JF (2004)
 Localized domains of G9a-mediated histone methylation are required for silencing of neuronal genes. Mol Cell 14:727–738
- Ballas N, Mandel G (2005) The many faces of REST oversee epigenetic programming of neuronal genes. Curr Opin Neurobiol 15:500–506
- Mehedint MG, Craciunescu CN, Zeisel SH (2010) Maternal dietary choline deficiency alters angiogenesis in fetal mouse hippocampus. Proc Natl Acad Sci USA 107:12834–12839
- Eichmann A, Corbel C, Le Douarin NM (1998) Segregation of the embryonic vascular and hemopoietic systems. Biochem Cell Biol 76:939–946
- Louissaint A Jr, Rao S, Leventhal C, Goldman SA (2002) Coordinated interaction of neurogenesis and angiogenesis in the adult songbird brain. Neuron 34:945–960
- 43. Munoz-Chapuli R, Perez-Pomares JM, Macias D, Garcia-Garrido L, Carmona R, Gonzalez M (1999) Differentiation of hemangio-blasts from embryonic mesothelial cells? A model on the origin of the vertebrate cardiovascular system. Differentiation 64:133–141
- Kinder SJ, Loebel DA, Tam PP (2001) Allocation and early differentiation of cardiovascular progenitors in the mouse embryo. Trends Cardiovasc Med 11:177–184
- 45. Risau W (1997) Mechanisms of angiogenesis. Nature 386:671–674
- 46. Zhu Y, Shwe Y, Du R, Chen Y, Shen FX, Young WL, Yang GY (2006) Effects of angiopoietin-1 on vascular endothelial growth factor-induced angiogenesis in the mouse brain. Acta Neurochir Suppl 96:438–443
- 47. Zacharek A, Chen J, Cui X, Li A, Li Y, Roberts C, Feng Y, Gao Q, Chopp M (2007) Angiopoietin1/Tie2 and VEGF/Flk1 induced

- by MSC treatment amplifies angiogenesis and vascular stabilization after stroke. J Cereb Blood Flow Metab 27:1684–1691
- Patan S (2004) Vasculogenesis and angiogenesis. Cancer Treat Res 117:3–32
- Kovacheva VP, Mellott TJ, Davison JM, Wagner N, Lopez-Coviella I, Schnitzler AC, Blusztajn JK (2007) Gestational choline deficiency causes global and Igf2 gene DNA hypermethylation by up-regulation of Dnmt1 expression. J Biol Chem 282:31777– 31788
- 50. Davison JM, Mellott TJ, Kovacheva VP, Blusztajn JK (2009) Gestational choline supply regulates methylation of histone H3, expression of histone methyltransferases G9a (Kmt1c) and Suv39h1 (Kmt1a), and DNA methylation of their genes in rat fetal liver and brain. J Biol Chem 284:1982–1989
- Garro AJ, McBeth DL, Lima V, Lieber CS (1991) Ethanol consumption inhibits fetal DNA methylation in mice: implications for the fetal alcohol syndrome. Alcohol Clin Exp Res 15:395–398
- Haycock PC, Ramsay M (2009) Exposure of mouse embryos to ethanol during preimplantation development: effect on DNA methylation in the h19 imprinting control region. Biol Reprod 81:618–627
- Bielawski DM, Zaher FM, Svinarich DM, Abel EL (2002) Paternal alcohol exposure affects sperm cytosine methyltransferase messenger RNA levels. Alcohol Clin Exp Res 26:347–351
- 54. Ouko LA, Shantikumar K, Knezovich J, Haycock P, Schnugh DJ, Ramsay M (2009) Effect of alcohol consumption on CpG methylation in the differentially methylated regions of H19 and IG-DMR in male gametes: implications for fetal alcohol spectrum disorders. Alcohol Clin Exp Res 33:1615–1627
- Kim Y-I, Miller JW, da Costa K-A, Nadeau M, Smith D, Selhub J, Zeisel SH, Mason JB (1994) Severe folate deficiency causes secondary depletion of choline and phosphocholine in liver. J Nutr 124:2197–2203
- Selhub J, Seyoum E, Pomfret EA, Zeisel SH (1991) Effects of choline deficiency and methotrexate treatment upon liver folate content and distribution. Cancer Res 51:16–21
- Varela-Moreiras G, Selhub J, da Costa K, Zeisel SH (1992) Effect of chronic choline deficiency in rats on liver folate content and distribution. J Nutr Biochem 3:519–522
- Barak AJ, Beckenhauer HC, Tuma DJ (1996) Betaine, ethanol, and the liver: a review. Alcohol 13:395–398
- Biller A, Bartsch AJ, Homola G, Solymosi L, Bendszus M (2009) The effect of ethanol on human brain metabolites longitudinally characterized by proton MR spectroscopy. J Cereb Blood Flow Metab 29:891–902
- Villanueva JA, Halsted CH (2004) Hepatic transmethylation reactions in micropigs with alcoholic liver disease. Hepatology 39:1303–1310
- Martinez-Lopez N, Varela-Rey M, Ariz U, Embade N, Vazquez-Chantada M, Fernandez-Ramos D, Gomez-Santos L, Lu SC, Mato JM, Martinez-Chantar ML (2008) S-adenosylmethionine and proliferation: new pathways, new targets. Biochem Soc Trans 36:848–852
- Manari AP, Preedy VR, Peters TJ (2003) Nutritional intake of hazardous drinkers and dependent alcoholics in the UK. Addict Biol 8:201–210
- Jiang R, Hu FB, Giovannucci EL, Rimm EB, Stampfer MJ, Spiegelman D, Rosner BA, Willett WC (2003) Joint association of alcohol and folate intake with risk of major chronic disease in women. Am J Epidemiol 158:760–771
- 64. Halsted CH, Robles EA, Mezey E (1973) Intestinal malabsorption in folate-deficient alcoholics. Gastroenterology 64:526–532
- 65. McMartin KE, Collins TD, Eisenga BH, Fortney T, Bates WR, Bairnsfather L (1989) Effects of chronic ethanol and diet treatment on urinary folate excretion and development of folate deficiency in the rat. J Nutr 119:1490–1497



- Ji C, Kaplowitz N (2003) Betaine decreases hyperhomocysteinemia, endoplasmic reticulum stress, and liver injury in alcohol-fed mice. Gastroenterology 124:1488–1499
- Barak AJ, Beckenhauer HC, Mailliard ME, Kharbanda KK, Tuma DJ (2003) Betaine lowers elevated s-adenosylhomocysteine levels in hepatocytes from ethanol-fed rats. J Nutr 133:2845–2848
- Craciunescu CN, Albright CD, Mar MH, Song J, Zeisel SH (2003) Choline availability during embryonic development alters progenitor cell mitosis in developing mouse hippocampus. J Nutr 133:3614

 –3618
- 69. Uban KA, Sliwowska JH, Lieblich S, Ellis LA, Yu WK, Weinberg J, Galea LA (2010) Prenatal alcohol exposure reduces the proportion of newly produced neurons and glia in the dentate gyrus of the hippocampus in female rats. Horm Behav 58:835–843
- Crews FT, Miller MW, Ma W, Nixon K, Zawada WM, Zakhari S (2003) Neural stem cells and alcohol. Alcohol Clin Exp Res 27:324–335
- Barnes DE, Walker DW (1981) Prenatal ethanol exposure permanently reduces the number of pyramidal neurons in rat hippocampus. Brain Res 227:333–340
- Hicks SD, Middleton FA, Miller MW (2010) Ethanol-induced methylation of cell cycle genes in neural stem cells. J Neurochem 114:1767–1780
- Thomas JD, Idrus NM, Monk BR, Dominguez HD (2010) Prenatal choline supplementation mitigates behavioral alterations associated with prenatal alcohol exposure in rats. Birth Defects Res A Clin Mol Teratol 88:827–837
- 74. Meck WH, Williams CL, Cermak JM, Blusztajn JK (2007) Developmental periods of choline sensitivity provide an ontogenetic mechanism for regulating memory capacity and age-related dementia. Front Integr Neurosci 1:7

- Moon J, Chen M, Gandhy SU, Strawderman M, Levitsky DA, Maclean KN, Strupp BJ (2010) Perinatal choline supplementation improves cognitive functioning and emotion regulation in the Ts65Dn mouse model of Down syndrome. Behav Neurosci 124:346–361
- Hagberg B, Aicardi J, Dias K, Ramos O (1983) A progressive syndrome of autism, dementia, ataxia, and loss of purposeful hand use in girls: Rett's syndrome: report of 35 cases. Ann Neurol 14:471–479
- 77. Jung BP, Jugloff DG, Zhang G, Logan R, Brown S, Eubanks JH (2003) The expression of methyl CpG binding factor MeCP2 correlates with cellular differentiation in the developing rat brain and in cultured cells. J Neurobiol 55:86–96
- Mill J, Petronis A (2008) Pre- and peri-natal environmental risks for attention-deficit hyperactivity disorder (ADHD): the potential role of epigenetic processes in mediating susceptibility. J Child Psychol Psychiatry 49:1020–1030
- Mill J, Tang T, Kaminsky Z, Khare T, Yazdanpanah S, Bouchard L, Jia P, Assadzadeh A, Flanagan J, Schumacher A, Wang SC, Petronis A (2008) Epigenomic profiling reveals DNA-methylation changes associated with major psychosis. Am J Hum Genet 82:696–711
- da Costa KA, Kozyreva OG, Song J, Galanko JA, Fischer LM, Zeisel SH (2006) Common genetic polymorphisms affect the human requirement for the nutrient choline. FASEB J 20:1336– 1344
- Kohlmeier M, da Costa KA, Fischer LM, Zeisel SH (2005) Genetic variation of folate-mediated one-carbon transfer pathway predicts susceptibility to choline deficiency in humans. Proc Natl Acad Sci USA 102:16025–16030

